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Minor dammarane saponins from the hongshen extract of Shenmai injection

Shi-Jin Qu^a, Jun-Jie Tan^a, Jian-Guo Cai^b, Yi-Ping Ling^b, Shan-Fei Zhang^b, Chang-Heng Tan^{a*} and Da-Yuan Zhu^a

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A new dammarane-type triterpenoid saponin, (20R)-ginsenoside ST₂ (1), along with five known saponins was isolated from the hongshen extract of Shenmai injection. The structure of 1 was elucidated to be (20R)-dammar-23(*E*)-ene-3 β ,6 α ,12 β ,20,25-pentol 6-*O*- β -D-glucopyranoside by means of spectroscopic methods.

Keywords: Shenmai injection; hongshen extract; dammarane saponins; (20R)-ginsenoside ST₂

1. Introduction

Shenmai injection (SMI) is a formula derived from the traditional Chinese medicine recipe Shen Mai San [1] and made of the extracts of hongshen (red ginseng, Radix Ginseng Rubra) and maidong [roots of Ophiopogon japonica (Thunb.) Ker-Gawl., Radix Ophiopogonis], which has been used to treat cardiac emergencies, such as coronary atherosclerotic cardiopathy, viral myocarditis, and myocardial infarction, and also to raise tumor patient's immunity. HPLC/MS analysis of SMI showed that its main ingredients were water-soluble ginsenosides from hongshen and a few from maidong [2,3]. In order to improve the quality control level of SMI, we conducted a chemical investigation to search the minor ingredients of the hongshen extract of SMI, which led us to the isolation of a new dammarane saponin, (20R)-ginsenoside ST₂ (1), along with five known triterpenoid glycosides, (20S)- ginsenoside ST_2 (2) [4], notoginsenosides R_8 (3) and R_9 (4) [5], notoginaxoside A (5) [6], and 25-hydroxy-(20*S*)-ginsenoside Rh_1 (6) [7] (Figure 1). The above ginsenosides are reported from hongshen for the first time.

2. Results and discussion

Compound **1**, a white amorphous powder, has the molecular formula of $C_{36}H_{62}O_{10}$ as deduced from the HR-ESI-MS (found $[M - H]^- m/z$ 653.4267, calcd 653.4265). The sugar moiety of **1** was elucidated to be a β -D-glucopyranosyl on the basis of the ¹H and ¹³C NMR signals at δ_H 5.06 (d, J = 7.6 Hz) and δ_C 106.1 (d), 79.8 (d), 78.3 (d), 75.6 (d), 72.0 (d), and 63.0 (t), as well as acid hydrolysis experiment and GC analysis. The ¹H and ¹³C NMR spectra (Table 1) of **1** for aglycone showed signals for a pair of *E*-double bond (δ_H 6.48 and 6.03, $J_{gemical} = 15.5$ Hz; δ_C 123.0 d and 142.6 d), three oxygenated methines (δ_C

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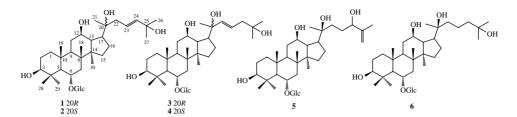


Figure 1. Structures of compounds 1-6.

80.2, 78.7, and 71.0) and two oxygenbearing quaternary carbons ($\delta_{\rm C}$ 74.0 and 70.0), as well as 23 saturated carbons (8 CH₃, 7 CH₂, 4 CH, and 4 C), indicating a dammarane-type triterpenoid [7]. The ^{13}C NMR spectral data of the aglycone of 1 were nearly superposed with those of (20R)-protopanaxatriol derivatives, such as 3 and (20R)-ginsenoside Rh₁ [7], except for the signals due to the side chain. In the side chain, two hydroxyls at C-20 and C-25, and the E-double bond between C-23 and C-24 were elucidated by the HMBC correlations (Figure 2). The stereochemistry of C-20 of 1 was determined to be Raccording to the ¹³C chemical shift of C-17 at δ 51.0, while that of 20S-epimer is usually observed at δ 54–55 as indicated by Teng et al. [7]. The above evidences demonstrated the structure of 1 to be (20R)-dammar-23(E)-ene-3 β ,6 α ,12 β ,20, 25-pentol 6-O-β-D-glucopyranoside.

Compound **2** was identified to be (20*S*)-epimer of **1**, i.e. ginsenoside ST₂, by comparison of its NMR, MS, and physical properties with those reported in the literature [4]. The major difference of the ¹³C NMR spectral data between (20*R*)- and (20*S*)-ginsenoside ST₂ (**1** and **2**) was observed at C-17, C-21, and C-22 ($\Delta \delta_{1-2} = -3.3$, -5.1, and +6.3 ppm, respectively).

Previous studies have disclosed that red ginseng (steamed ginseng) had many less polar dammarane saponins with various side chains that were not present in white ginseng due to the high temperature during the preparation procedure [8]. Compounds 1-6 might be related with the special process of the hongshen extract.

3. Experimental

3.1 General experimental procedures

Optical rotations were determined on a Perkin-Elmer 341 polarimeter. The IR spectra were recorded on a Nicolet-Magna-750-FTIR spectrometer. The NMR spectra were taken on a Bruker AV-400 spectrometer with TMS as internal standard. ESI-MS and HR-ESI-MS were obtained on an Esquire 3000plus and a Q-TOF-Ultima mass spectrometers, respectively. Silica gel (200-300 mesh, Qingdao Haiyang Chemical Co. Ltd., Qingdao, China), D-1400 macroporous resin (Yangzhou Pharmaceutical Factory, Yangzhou, China), and C₁₈-reversed phase silica gel (150-200 mesh, Fuji Silysia Chemical Ltd, Aichi, Japan) were used for column chromatography (CC). Silica gel HSGF₂₅₄ (Yantai Jiangyou Guijiao Kaifa Co., Yantai, China) was used for TLC. GC analysis was performed on a Perkin-Elmer Sigma-115 gas chromatograph. Analytical and prepared HPLC isolation were conducted on a Varian HPLC system (pump: Prepstar SD-1, detector: UVvis 320, column: Merck, 5 µm, I.D. 4.6×250 mm, and Merck 12 µm, I.D. 25×250 mm, respectively).

3.2 Plant material

The hongshen extract was prepared as per the manufacture procedure of SMI, provided by the Chiatai Qingchunbao Phar-

Site	δ_{C} of 1	$\delta_{ m H}$ of 1	$\delta_{\rm C}$ of ${f 2}$	$\delta_{ m H}$ of ${f 2}$
1	39.6 t	1.69 (dd, 12.7, 6.8), 1.03 (m)	39.6 t	1.76 (dd, 11.4, 4.8), 1.06 (m)
2	28.1 t	1.89 (m), 1.83 (m)	28.1 t	1.94 (m), 1.87 (m)
3	78.7 d	3.53 (dd, 11.0, 5.2)	78.8 d	3.56 (dd, 11.0, 5.3)
4	40.5 s	_	40.5 s	_
5	61.5 d	1.46 (d, 10.7)	61.6 d	1.47 (d, 10.5)
6	80.2 d	4.47 (td, 10.8, 2.0)	80.2 d	4.47 (td, 10.5, 2.5)
7	45.3 t	2.56 (dd, 12.3, 2.0),	45.4 t	2.56 (dd, 12.5, 2.5),
		1.90 (dd, 12.3, 10.8)		1.98 (dd, 12.5, 10.5)
8	41.2 s	_	41.3 s	_
9	50.3 d	1.59 (m)	50.4 d	1.59 (m)
10	39.8 s	_	39.8 s	_
11	32.3 t	2.18 (m), 1.59 (m)	32.3 t	2.17 (m), 1.59 (m)
12	71.0 d	3.90 (m)	71.2 d	3.91 (m)
13	49.2 d	2.04 (m)	48.7 d	2.12 (m)
14	51.8 s	_	51.8 s	_
15	31.4 t	1.70 (dd, 10.8, 3.0),	31.4 t	1.70 (dd, 12.0, 3.8),
		1.16 (br t, 10.1)		1.16 (t, 12.0)
16	26.5 t	1.97 (m), 1.33 (m)	26.9 t	1.81 (m), 1.40 (m)
17	51.0 d	2.38 (m)	54.3 d	2.32 (m)
18	17.6 q	1.26 (s, 3H)	17.6	1.29 (s, 3H)
19	17.8 q	1.08 (s, 3H)	17.8	1.10 (s, 3H)
20	74.0 s	_	73.5	_
21	22.6 q	1.43 (s, 3H)	27.7 q	1.42 (s, 3H)
22	46.3 t	2.54 (dd, 13.9, 6.9),	40.0 t	2.83 (dd, 13.7, 5.3),
		2.44 (dd, 13.9, 7.8)		2.48 (dd, 13.7, 8.9)
23	123.0 d	6.48 (ddd, 15.5, 7.8, 6.9)	123.4 d	6.34 (ddd, 15.6, 8.9, 5.3)
24	142.6 d	6.03 (d, 15.5)	142.2 d	6.06 (d, 15.6)
25	70.0 s	_	70.0 s	_
26	30.9 q	1.55 (s, 3H)	30.9 q	1.57 (s, 3H)
27	30.9 q	1.55 (s, 3H)	30.9 q	1.56 (s, 3H)
28	31.9 q	2.08 (s, 3H)	31.9 q	2.09 (s, 3H)
29	16.5 q	1.63 (s, 3H)	16.5 q	1.63 (s, 3H)
30	17.1 q	0.84 (s, 3H)	17.0 q	0.86 (s, 3H)
Glu-1'	106.1 d	5.06 (d, 7.6)	106.1 d	5.06 (d, 7.5)
2'	75.6 d	4.12 (dd, 8.1, 7.8)	75.6 d	4.12 (t, 7.8)
3'	79.8 d	4.29 (t, 8.7)	79.8 d	4.28 (t, 8.7)
4'	72.0 d	4.24 (t, 8.8)	72.0 d	4.23 (t, 9.0)
5'	78.3 d	4.00 (m)	78.3 d	3.95 (m)
6'	63.0 t	4.57 (dd, 11.3, 1.4),	63.2 t	4.55 (dd, 11.2, 1.4),
		4.40 (dd, 11.3, 5.4)		4.37 (dd, 11.2, 5.4)

Table 1. 1 H and 13 C NMR spectral data of 1 and 2 (400 and 100 MHz, C₅D₅N).

maceutical Co. In brief, 10 kg of dried red ginseng (purchased from the ginseng market in Jilin Province) was refluxed 2 h in 30 liters of 80% EtOH. The extract was concentrated to 5 liters under reduced pressure, following that placed into refrigerator overnight, then the solid residue was filtered out. To the solution, 30 g of active carbon was added, stirred fully, and filtered after 30 min. The percolate was adjusted to pH 7.5-7.7 using 1% NaOH, then heated and kept boiling for 40 min, and decolorised again using 25 g of active carbon as in the above method. To the filtrate, distilled water was added to 5 liters to yield the hongshen extract.

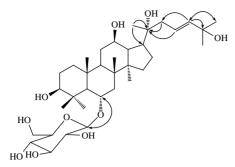


Figure 2. Selected HMBC correlations of 1.

3.3 Extraction and isolation

concentrated hongshen extract The (1 liters) was subjected to the CC of macroporous resin eluted with gradient EtOH in H₂O (0, 20%, 40%, 70%, 95%). The combined 40% and 70% EtOH fractions (about 13g) were subjected to CC of silica gel with gradient CHCl₃-MeOH as eluant. The CHCl₃-MeOH (6:1) fraction (about 1 g) was separated into subfractions 1-6 by CC of Rp-18 silica gel (increasing MeOH in H_2O , 25–60%). Fraction 3 (40% MeOH $-H_2O$, 150 mg) was further isolated by prepared HPLC system (CH₃CN-H₂O, 19%, UV, 202 nm) to obtain 1 (8 mg), 2 (14 mg), 3 (7 mg), 4 (15 mg), 5 (10 mg), and 6 (35 mg).

3.3.1 (20R)-Ginsenoside ST_2 (1)

White amorphous powder. $[\alpha]_D^{14} + 34$ (*c* 0.085, MeOH). IR (KBr) ν_{max} : 3421, 2926, 1630, 1462, 1385, 1155, 1078, 1028 cm⁻¹; ¹H and ¹³C NMR spectral data, see Table 1. ESI-MS *m/z* 653 [M - H]⁻, 699 [M + HCOO]⁻ (negative), 677 [M + Na]⁺ (positive); HR-ESI-MS (negative) *m/z* 653.4267 [M - H]⁻ (calcd for C₃₆H₆₁O₁₀, 653.4265).

3.3.2 Acid hydrolysis of 1

Compound 1 (1 mg) was refluxed in 2 N HCl-dioxane (1:1 v/v, 2 ml) for 2 h. On cooling, the mixture was neutralized with NaHCO₃. After extraction with AcOEt,

the aqueous layer was concentrated by blowing with N₂. The residue was purified by CC of Sephadex LH-20 (MeOH-H₂O 1:1, v/v) to give the sugar moiety. The purified sugar and standard D-glucose (Sigma-Aldrich, St Louis, USA) were treated with L-cysteine methyl ester hydrochloride (2 mg) in pyridine (1 ml) at 60°C for 1 h. Then, the solution was treated with N,O-bis(trimethylsilyl)trifluoroacetamide (0.02 ml) at 60°C for 1 h. The supernatant was subjected to GC analysis to identify the sugars. Conditions for GC were: capillary column, DB5-MS $(30 \text{ m} \times 0.25 \text{ mm} \times 0.25 \text{ }\mu\text{m})$, oven temperature program, 180-300°C at 6°C/min; injection temperature 350°C; carrier gas, He at 1 ml/min. D-Glucose was detected by comparing its retention time with that of the authentic sample ($t_{\rm R} = 12.28$ min).

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